

**VOLTage Fast Electrophoresis Buffer (20x)**

#GB02.0120 (1L 20X) | GB02.0520 (5L 20X) | GB02s (trial size)

(FOR RESEARCH ONLY)



**Product:** VOLTage Fast Electrophoresis Buffer is ideal for agarose gel electrophoresis of small DNA fragments (up to 1000bp). Unlike gels prepared with TAE or TBE, which tolerate only about 110V (mini-gel), this low-conductance electrophoretic buffer allows for the application of high voltage (200V-300V for a mini-gel) without generating a lot of heat, and without melting your gel. This allows for much faster runs. Not only does this save time, but faster runs result in less fuzzy bands, and therefore, using VOLTage Fast Electrophoresis Buffer results in very sharp bands. Moreover, in comparison with TBE and TAE, using VOLTage Fast Electrophoresis Buffer results in larger differences in relative mobility (Rf), especially between smaller DNA fragments, allowing for a better separation of DNA bands of similar size. On the other hand, if you already get the resolution you need, with VOLTage Fast Electrophoresis Buffer you can achieve the same result using less agarose, thus not only saving you time but money as well. Finally, gels prepared with VOLTage Fast Electrophoresis Buffer are clearer than gels prepared with TAE, increasing detection sensitivity, and also stronger, making them easier to handle.

**Applications:** Fast Electrophoresis of small DNA fragments (up to 1000bp)

**Contents:** #GB02.0120 contains 1L of 20X concentrated VOLTage Fast Electrophoresis Buffer  
#GB02.0520 contains 5L of 20X concentrated VOLTage Fast Electrophoresis Buffer  
# GB02s is a trial sample (100ml) of 20X concentrated VOLTage Fast Electrophoresis Buffer

**Properties:** Time Saving  
High Resolution  
Better Separation  
Clearer and Stronger Gels

**Storage:** Store at room temperature for up to 3 years.

### Prior to use:

VOLTage Fast Electrophoresis Buffer is prepared with ultrapure water and 0.2µm filtered, and is provided as a 20X concentrated aqueous solution. The working concentration is 1X. Prepare 1L VOLTage Fast Electrophoresis Buffer by mixing 50ml of the 20x concentrate with 950ml of ddH<sub>2</sub>O. VOLTage Fast Electrophoresis Buffer (1X) should be used both for the preparation of the agarose gel as well as the running buffer. Like TBE (and unlike TAE), VOLTage Fast Electrophoresis Buffer does not exhaust quickly during extended electrophoresis, and so the buffer may be used multiple times.

### Usage

Prepare 1X VOLTage Fast Electrophoresis Buffer by diluting the concentrate in pure water. Prepare the agarose gel with 1X VOLTage Fast Electrophoresis Buffer as usual (like with TAE or TBE). Fill the electrophoresis chambers with 1X VOLTage Fast Electrophoresis Buffer and put the gel casting tray into place. It is recommended to wash the electrophoresis chambers the first time with VOLTage Fast Electrophoresis Buffer to remove any residual TAE or TBE (if applicable). For optimal resolution, cover the gel with no more than 3-5mm of VOLTage Fast Electrophoresis Buffer.

Apply samples and run the gel at constant voltage, using a higher voltage than normal. GRiSP suggests to start by doubling the normal voltage one normally uses for either TAE or TBE and then gradually increase voltage to find the optimum. The optimal voltage depends, among others, on the type and percentage of agarose, as well as on the nature of the sample. Therefore, occasionally the optimal voltage might be less than 2x the normal voltage. Too high voltage does not result in melting of the gel, however, DNA bands might become distorted. The run time will be reduced considerably and will be roughly reduced by the same value as the increase of the voltage.

For example, when normally running a gel for at 100V for 1 hour, start with running at 200V for 30 minutes. Then, determine whether the electrophoretic conditions might be further improved (voltage, time, percentage agarose). It is strongly advised to determine optimum run time empirically, and not to rely on the tracking dye, as the behavior of tracking dyes depends highly on the electrophoresis buffer and the percentage of agarose (see section about tracking dyes hereunder).

### Tracking dyes

Nucleic acid agarose gel electrophoresis loading buffers contain one or more tracking dyes in order to help with pipetting and to be able to visually follow the electrophoresis progress. Common dyes are Bromophenol blue (BPB), Xylene Cyanol FF (XC), Orange G (OG), and Cresol Red (CR). Migration of these dyes depends, among other factors, on the electrophoresis buffer used. In general, when using VOLTage Fast Electrophoresis Buffer, tracking dyes migrate much faster than with either TAE or TBE, that is: co-migrate with smaller DNA fragments than usual. The following table compares the approximate migration rates of tracking dyes using various concentrations of a standard agarose in TAE, TBE, and VOLTage Fast Electrophoresis Buffer. Note that other factors, such as the type of agarose and the DNA sample, are also of influence and that, therefore, these values should only be used as a rough estimation.

% agarose	Xylene Cyanol FF			Cresol Red	Bromophenol Blue			Orange G
	VOLTage Fast	TAE	TBE	VOLTage Fast	VOLTage Fast	TAE	TBE	VOLTage Fast
<b>0.25/0.30*</b>	> 25 kb	25 kb	19 kb	2.5-3.0 kb	1.5 kb	2.9 kb	2.9 kb	400 bp
<b>0.50</b>	15 kb	11 kb	12 kb	750 bp	400 bp	1.7 kb	1.4 kb	60 bp
<b>0.75</b>	12 kb	10 kb	9 kb	325 bp	200 bp	1.0 kb	700 bp	50 bp
<b>1.00</b>	10 kb	6 kb	4 kb	250 bp	125 bp	500 bp	400 bp	<40 bp
<b>1.25</b>	2.1 kb	3.6 kb	2.5 kb	170 bp	90 bp	370 bp	250 bp	<40 bp
<b>1.50</b>	900 bp	2.8 kb	1.8 kb	100 bp	50 bp	300 bp	200 bp	< 40 bp
<b>1.75</b>	n.d.	1.8 kb	1.1 kb	n.d.	n.d.	200 bp	100 bp	n.d.
<b>2.00</b>	350 bp	1.3 kb	850 bp	50 bp	<40 bp	150 bp	70 bp	< 40 bp
<b>2.50</b>	250 bp	n.d.	n.d.	< 40 bp	< 40 bp	n.d.	n.d.	< 40 bp
<b>3.00</b>	150 bp	n.d.	n.d.	< 40 bp	< 40 bp	n.d.	n.d.	< 40 bp

\*) for TAE/TBE: 0.30% and for VOLTage Fast: 0.25%; Note that bp sizes were rounded off. Data for TAE and TBE were compiled from data provided by the agarose manufacturer and thirds, and were partly confirmed by GRiSP